



## Product Information

### T4 DNA Polymerase

Part Number:	P708L
Lot Number:	

Certificate of Analysis

2010-P708L-Rev.D

### Product Description:

T4 DNA Polymerase catalyzes the extension of a primed DNA template in the 5'→3' direction. This enzyme exhibits a powerful 3'→5' exonuclease activity, while lacking any inherent 5'→3' exonuclease or strand displacement functions.

### Source of Protein

Purified from a strain of *E. coli* that expresses the recombinant T4 DNA Polymerase gene.

### Supplied in

100 mM KPO<sub>4</sub>  
1.0 mM dithiothreitol  
0.1 mM EDTA  
50% glycerol  
pH 6.5 @ 25°C

### Supplied With

B011 (10X Blue Buffer)

### 10X Blue Buffer (B011)

500 mM NaCl  
100 mM Tris-HCl  
100 mM MgCl<sub>2</sub>  
10 mM DTT  
pH 7.9 @ 25°C

### Unit Definition

One unit is defined as the amount of enzyme that will incorporate 10 nmol of dNTP into acid-precipitable material in 30 minutes at 37°C.

## Product Specification\*

Unit Size:	2,000 Units
Unit Concentration	3,000 U/mL
Protein Concentration	0.54 mg/mL
Purity (SDS-PAGE)	>99%
Specific Activity	5,555 U/mg
SS Exonuclease	Functional
DS Exonuclease	Functional
Endonuclease	30 U <10% converted
<i>E. coli</i> 16S rDNA Contamination	30 U <10 copies
Storage	-20°C

\* For a detailed summary of assay conditions and data, refer to the Quality Controls Analysis section below

### Quality Control Analysis:

#### Unit Characterization Assay

Unit activity was measured using a 2-fold serial dilution method. Dilutions of enzyme were made in 1X reaction buffer ([T4 DNA Polymerase]<sub>f</sub> = 0.25-0.002µg/µL) and added to 50 µL reactions containing 10µg denatured Calf Thymus DNA, 1X Blue Buffer, 4mCi/mL <sup>3</sup>H-dTTP and 100 µM dNTPs. Reactions were incubated 10 minutes at 37°C, plunged on ice, and analyzed using the method of Sambrook and Russell (*Molecular Cloning, v3, 2001, pp. A8.25-A8.26*)

#### Protein Concentration (OD<sub>280</sub>) Measurement

A 2.0 µL sample of enzyme was analyzed at OD<sub>280</sub> using a Nanodrop ND-1000 spectrophotometer standardized using a 2.0 mg/ml BSA sample (Pierce Cat #23209) and blanked with product storage solution. The observed average measurement of 3 replicate samples was converted to mg/mL using an extinction coefficient of 128,440 and molecular weight of 103,609 Daltons. Acceptance for this assay is +/- 5% of reference sample.

#### SDS-Page (Physical Purity Assessment)

2.0 µL of enzyme solution was loaded on a denaturing 4-20% Tris-Glycine SDS-PAGE gel flanked by a broad-range MW marker and 2.0 µL of a 1:100 dilution of the sample. Following electrophoresis, the gel was stained and the samples compared to determine physical purity. The acceptance criteria for this test requires that the aggregate mass of contaminant bands in the concentrated sample do not exceed the mass of the protein of interest band in the dilute sample, confirming greater than 99% purity of the concentrated sample.

## **Functional Tests:**

### **Single-Stranded Exonuclease Activity**

A 50  $\mu$ L reaction containing 11,000 cpm of a radiolabeled single-stranded DNA substrate and 10  $\mu$ L of enzyme solution incubated for 4 hours at 37°C resulted in greater than 80% release of TCA-soluble counts.

### **Double-Stranded Exonuclease Activity**

A 50  $\mu$ L reaction containing 5,000 cpm of a radiolabeled double-stranded DNA substrate and 10  $\mu$ L of enzyme solution incubated for 4 hours at 37°C resulted in greater than 50% release of TCA-soluble counts.

## **Nuclease Contamination Tests:**

### **Endonuclease Activity**

A 50  $\mu$ L reaction containing 0.5  $\mu$ g of pBR322 DNA and 10  $\mu$ L of enzyme solution incubated for 4 hours at 37°C resulted in no visually discernible conversion to nicked circular DNA as determined by agarose gel electrophoresis.

### ***E. coli* 16S rDNA Contamination Test**

Replicate 5  $\mu$ L samples of enzyme solution were denatured and screened in a TaqMan qPCR assay for the presence of contaminating *E. coli* genomic DNA using oligonucleotide primers corresponding to the 16S rRNA locus. The acceptance criterion for the test is the threshold cycle count ( $C_t$ ) produced by the average of 3 replicate no template control samples. Based on the correlation between the no template control  $C_t$  values, and standard curve data, the detection limit of this assay is <10 copies genome/sample.

## **References**

1. Tabor, S. and Struhl, K. (1989) In DNA-Dependent DNA Polymerases. F. M. Ausebel, R. Brent, R. E. Kingston, D. D. Moore, J. G. Seidman, J. A. Smith and K. Struhl (Eds.), Current Protocols in Molecular Biology, pp. 3.5.10-3.5.12.
2. Sambrook, J., Fritsch, E.F. and Maniatis, T. (1989) Molecular Cloning: A Laboratory Manual, (2nd Ed.), 5.44-5.47.
2. Dale, R., McClure, B. and Houchins, J. (1985) Plasmid, 13, 31-40.
3. Kunkel, T.A., Roberts, J.D. and Zakour, R.A. (1987) R. Wu and L. Grossman (Eds.), Methods Enzymol., 154, pp. 367-382. San Diego: Academic Press.
4. Panet, A., van de Sande, J.H., Loewen, P.C. and Khorana, H.G. (1973) Biochemistry, 12, 5045-5050.



### **Limitations of Use**

This product was developed, manufactured, and sold for *in vitro* use only. The product is not suitable for administration to humans or animals. MSDS sheets relevant to this product are available upon request.